

Quantifiler Trio DNA Quantification - Sample Sheet

#	Sample Name	Well	Comments
1	Trio Standard 1	A1	
2	Trio Standard 2	B1	
3	Trio Standard 3	C1	
4	Trio Standard 4	D1	
5	Trio Standard 5	E1	
6	NTC	F1	
7		G1	
8		H1	
9	Trio Standard 1	A2	
10	Trio Standard 2	B2	
11	Trio Standard 3	C2	
12	Trio Standard 4	D2	
13	Trio Standard 5	E2	
14	NTC	F2	
15		G2	
16		H2	
17		A3	
18		B3	
19		C3	
20		D3	
21		E3	
22		F3	
23		G3	
24		H3	
25		A4	
26		B4	
27		C4	
28		D4	
29		E4	
30		F4	
31		G4	
32		H4	
33		A5	
34		B5	
35		C5	
36		D5	
37		E5	
38		F5	
39		G5	
40		H5	
41		A6	
42		B6	
43		C6	
44		D6	
45		E6	
46		F6	
47		G6	
48		H6	
49		A7	
50		B7	
51		C7	
52		D7	
53		E7	
54		F7	
55		G7	
56		H7	
57		A8	
58		B8	
59		C8	
60		D8	
61		E8	
62		F8	
63		G8	
64		H8	
65		A9	
66		B9	
67		C9	
68		D9	

#	Sample Name	Well	Comments
69		E9	
70		F9	
71		G9	
72		H9	
73		A10	
74		B10	
75		C10	
76		D10	
77		E10	
78		F10	
79		G10	
80		H10	
81		A11	
82		B11	
83		C11	
84		D11	
85		E11	
86		F11	
87		G11	
88		H11	
89		A12	
90		B12	
91		C12	
92		D12	
93		E12	
94		F12	
95		G12	
96		H12	

Date:

epMotion Quantitation Plate Set-up

	1	2	3	4	5	6	7	8	9	10	11	12
A	A_50ng Standard	50ng Standard	0	0	0	0	0	0	0	0	0	0
B	A_5ng Standard	5ng Standard	0	0	0	0	0	0	0	0	0	0
C	A_5ng Standard	5ng Standard	0	0	0	0	0	0	0	0	0	0
D	A_0.05ng Standard	0.05ng Standard	0	0	0	0	0	0	0	0	0	0
E	A_0.005ng Standard	0.005ng Standard	0	0	0	0	0	0	0	0	0	0
F	A_NTC	NTC	0	0	0	0	0	0	0	0	0	0
G	0	0	0	0	0	0	0	0	0	0	0	0
H	0	0	0	0	0	0	0	0	0	0	0	0

Run ID: _____

7500: _____

Quantitation performed by: _____

epMotion setup check: _____

	Kit Component	Volume per reaction (µl)	# of sample (+ extra)	Amount to pipette (µl)
Trio Master Mix		10		0
Trio Primer / IPC Mix		8		0

B2: Samples 1-24

	1	2	3	4	5	6
A						
B						
C						
D						

B3: Samples 25-48

	1	2	3	4	5	6
A						
B						
C						
D						

C3: Samples 49-72

	1	2	3	4	5	6
A						
B						
C						
D						

C4: Samples 73-96

	1	2	3	4	5	6
A						
B						
C						
D						

Quantifiler Trio DNA Quantification - Plate Map

[illegible]

Quantifiler Trio DNA Quantification - Report

Plate:	
Date:	
Analyst:	

Well	Sample Name	M:F Ratio	D.I.	IPC C _t	Small Auto	Y (male)	Large Auto
A1	Trio Standard 1						
B1	Trio Standard 2						
C1	Trio Standard 3						
D1	Trio Standard 4						
E1	Trio Standard 5						
F1	NTC						
G1							
H1							
A2	Trio Standard 1						
B2	Trio Standard 2						
C2	Trio Standard 3						
D2	Trio Standard 4						
E2	Trio Standard 5						
F2	NTC						
G2							
H2							
A3							
B3							
C3							
D3							
E3							
F3							
G3							
H3							
A4							
B4							
C4							
D4							
E4							
F4							
G4							
H4							
A5							
B5							
C5							
D5							
E5							
F5							
G5							
H5							
A6							
B6							
C6							
D6							
E6							
F6							
G6							
H6							
A7							
B7							
C7							
D7							
E7							
F7							
G7							
H7							
A8							
B8							
C8							
D8							
E8							
F8							
G8							
H8							
A9							
B9							
C9							
D9							

Quantifiler Trio DNA Quantification - Report

Plate:	
Date:	
Analyst:	

Well	Sample Name	M:F Ratio	D.I.	IPC C _t	Small Auto	Y (male)	Large Auto
E9							
F9							
G9							
H9							
A10							
B10							
C10							
D10							
E10							
F10							
G10							
H10							
A11							
B11							
C11							
D11							
E11							
F11							
G11							
H11							
A12							
B12							
C12							
D12							
E12							
F12							
G12							
H12							

Post Microcon Concentration Calculations

Lot:[illegible]

Date:

[illegible]

Amplification Calculations

Date: _____

GlobalFiler

Kit Component		Volume per reaction (µl)	
GlobalFiler Master Mix		7.5	X
GlobalFiler Primer Mix		2.5	X

Y Filer

Kit Component		Volume per reaction (µl)	
PCR Reaction Mix		9.2	X
AmpFISTR Yfiler Primer Set		5	X
AmpliTaq Gold DNA Polymerase		0.8	X

# of samples (+ extra)	Amount to pipette
	0
	0

# of samples (+ extra)	Amount to pipette
	0
	0
	0

GlobalFiler

Amplification Date:_____

RX#	Sample	Conc ng/ul	Dil	Vol. Ampl. (ul)	TE	Results OK	RI
1				#DIV/0!	#DIV/0!		
2				#DIV/0!	#DIV/0!		
3				#DIV/0!	#DIV/0!		
4				#DIV/0!	#DIV/0!		
5				#DIV/0!	#DIV/0!		
6				#DIV/0!	#DIV/0!		
7				#DIV/0!	#DIV/0!		
8				#DIV/0!	#DIV/0!		
9				#DIV/0!	#DIV/0!		
10				#DIV/0!	#DIV/0!		
11				#DIV/0!	#DIV/0!		
12				#DIV/0!	#DIV/0!		
13				#DIV/0!	#DIV/0!		
14				#DIV/0!	#DIV/0!		
15				#DIV/0!	#DIV/0!		
16				#DIV/0!	#DIV/0!		
17				#DIV/0!	#DIV/0!		
18				#DIV/0!	#DIV/0!		
19				#DIV/0!	#DIV/0!		
20				#DIV/0!	#DIV/0!		
21				#DIV/0!	#DIV/0!		
22				#DIV/0!	#DIV/0!		
23				#DIV/0!	#DIV/0!		
24				#DIV/0!	#DIV/0!		
25				#DIV/0!	#DIV/0!		
26				#DIV/0!	#DIV/0!		
27				#DIV/0!	#DIV/0!		
28				#DIV/0!	#DIV/0!		
29				#DIV/0!	#DIV/0!		
30				#DIV/0!	#DIV/0!		
31				#DIV/0!	#DIV/0!		
32				#DIV/0!	#DIV/0!		
33				#DIV/0!	#DIV/0!		
34				#DIV/0!	#DIV/0!		
35				#DIV/0!	#DIV/0!		
36				#DIV/0!	#DIV/0!		
37				#DIV/0!	#DIV/0!		
38				#DIV/0!	#DIV/0!		
39				#DIV/0!	#DIV/0!		
40				#DIV/0!	#DIV/0!		

Amplification performed by: _____
 Work area decontaminated before and after use _____
 Run temp verified _____
 Pre-amp tech review performed by/date: _____

Thern
Order
Instru

GlobalFiler

RX#	Sample	Conc ng/ul	Dil	Vol. Ampl. (ul)	TE	Results OK	RI
41				#DIV/0!	#DIV/0!		
42				#DIV/0!	#DIV/0!		
43				#DIV/0!	#DIV/0!		
44				#DIV/0!	#DIV/0!		
45				#DIV/0!	#DIV/0!		
46				#DIV/0!	#DIV/0!		
47				#DIV/0!	#DIV/0!		
48				#DIV/0!	#DIV/0!		
49				#DIV/0!	#DIV/0!		
50				#DIV/0!	#DIV/0!		
51				#DIV/0!	#DIV/0!		
52				#DIV/0!	#DIV/0!		
53				#DIV/0!	#DIV/0!		
54				#DIV/0!	#DIV/0!		
55				#DIV/0!	#DIV/0!		
56				#DIV/0!	#DIV/0!		
57				#DIV/0!	#DIV/0!		
58				#DIV/0!	#DIV/0!		
59				#DIV/0!	#DIV/0!		
60				#DIV/0!	#DIV/0!		
61				#DIV/0!	#DIV/0!		
62				#DIV/0!	#DIV/0!		
63				#DIV/0!	#DIV/0!		
64				#DIV/0!	#DIV/0!		
65				#DIV/0!	#DIV/0!		
66				#DIV/0!	#DIV/0!		
67				#DIV/0!	#DIV/0!		
68				#DIV/0!	#DIV/0!		
69				#DIV/0!	#DIV/0!		
70				#DIV/0!	#DIV/0!		
71				#DIV/0!	#DIV/0!		
72				#DIV/0!	#DIV/0!		
73				#DIV/0!	#DIV/0!		
74				#DIV/0!	#DIV/0!		
75				#DIV/0!	#DIV/0!		
76				#DIV/0!	#DIV/0!		
77				#DIV/0!	#DIV/0!		
78				#DIV/0!	#DIV/0!		
79				#DIV/0!	#DIV/0!		
80				#DIV/0!	#DIV/0!		

GlobalFiler

RX#	Sample	Conc ng/ul	Dil	Vol. Ampl. (ul)	TE	Results OK	RI
81				#DIV/0!	#DIV/0!		
82				#DIV/0!	#DIV/0!		
83				#DIV/0!	#DIV/0!		
84				#DIV/0!	#DIV/0!		
85				#DIV/0!	#DIV/0!		
86				#DIV/0!	#DIV/0!		
87				#DIV/0!	#DIV/0!		
88				#DIV/0!	#DIV/0!		
89				#DIV/0!	#DIV/0!		
90				#DIV/0!	#DIV/0!		
91				#DIV/0!	#DIV/0!		
92				#DIV/0!	#DIV/0!		
93				#DIV/0!	#DIV/0!		
94				#DIV/0!	#DIV/0!		
95				#DIV/0!	#DIV/0!		
96				#DIV/0!	#DIV/0!		

[illegible]

nal cyler:_____

verified: _____

ment_____

[illegible]

[illegible]

GlobalFiler

1 of 2

Amplification Date: _____

RX#	Sample	Results OK	RI	RI 8 sec	RI 3 sec	Amp More	Amp Less	Comments
1								
2								
3								
4								
5								
6								
7								
8								
9								
10								
11								
12								
13								
14								
15								
16								
17								
18								
19								
20								
21								
22								
23								
24								
25								
26								
27								
28								
29								
30								
31								
32								
33								
34								
35								
36								
37								
38								
39								
40								

Amplification performed by: _____
Work area decontaminated before and after use: _____
Pre-amp tech review performed by/date: _____
epMotion setup check: _____

Thermal cycler: _____
Instrument: _____
Run temp verified: _____

GlobalFiler

2 of 2

RX#	Sample	Results OK	RI	RI 8 sec	RI 3 sec	Amp More	Amp Less	Comments
41								
42								
43								
44								
45								
46								
47								
48								
49								
50								
51								
52								
53								
54								
55								
56								
57								
58								
59								
60								
61								
62								
63								
64								
65								
66								
67								
68								
69								
70								
71								
72								
73								
74								
75								
76								
77								
78								
79								
80								

GlobalFiler

3 of 3

RX#	Sample	Results OK	RI	RI 8 sec	RI 3 sec	Amp More	Amp Less	Comments
81								
82								
83								
84								
85								
86								
87								
88								
89								
90								
91								
92								
93								
94								
95								
96								

Yfiler

Amplification Date: _____

RX#	Sample	Conc ng/ul	Dil	Vol. Ampl. (ul)	TE	Results OK	RI	RI 8 sec	RI 3 sec
1				#DIV/0!	#DIV/0!				
2				#DIV/0!	#DIV/0!				
3				#DIV/0!	#DIV/0!				
4				#DIV/0!	#DIV/0!				
5				#DIV/0!	#DIV/0!				
6				#DIV/0!	#DIV/0!				
7				#DIV/0!	#DIV/0!				
8				#DIV/0!	#DIV/0!				
9				#DIV/0!	#DIV/0!				
10				#DIV/0!	#DIV/0!				
11				#DIV/0!	#DIV/0!				
12				#DIV/0!	#DIV/0!				
13				#DIV/0!	#DIV/0!				
14				#DIV/0!	#DIV/0!				
15				#DIV/0!	#DIV/0!				
16				#DIV/0!	#DIV/0!				
17				#DIV/0!	#DIV/0!				
18				#DIV/0!	#DIV/0!				
19				#DIV/0!	#DIV/0!				
20				#DIV/0!	#DIV/0!				
21				#DIV/0!	#DIV/0!				
22				#DIV/0!	#DIV/0!				
23				#DIV/0!	#DIV/0!				
24				#DIV/0!	#DIV/0!				
25				#DIV/0!	#DIV/0!				
26				#DIV/0!	#DIV/0!				
27				#DIV/0!	#DIV/0!				
28				#DIV/0!	#DIV/0!				
29				#DIV/0!	#DIV/0!				
30				#DIV/0!	#DIV/0!				
31				#DIV/0!	#DIV/0!				
32				#DIV/0!	#DIV/0!				
33				#DIV/0!	#DIV/0!				
34				#DIV/0!	#DIV/0!				
35				#DIV/0!	#DIV/0!				
36				#DIV/0!	#DIV/0!				
37				#DIV/0!	#DIV/0!				
38				#DIV/0!	#DIV/0!				
39				#DIV/0!	#DIV/0!				
40				#DIV/0!	#DIV/0!				

Work area decontaminated before and after use: _____

Run Temp Verified: _____

Thermal Cycler: _____

Pre-amp tech review performed by/date: _____

Order Verified: _____

Instrument: _____

Amplification performed by: _____

Yfiler

RX#	Sample	Conc ng/ul	Dil	Vol. Ampl. (ul)	TE	Results OK	RI	RI 8 sec	RI 3 sec
41				#DIV/0!	#DIV/0!				
42				#DIV/0!	#DIV/0!				
43				#DIV/0!	#DIV/0!				
44				#DIV/0!	#DIV/0!				
45				#DIV/0!	#DIV/0!				
46				#DIV/0!	#DIV/0!				
47				#DIV/0!	#DIV/0!				
48				#DIV/0!	#DIV/0!				
49				#DIV/0!	#DIV/0!				
50				#DIV/0!	#DIV/0!				
51				#DIV/0!	#DIV/0!				
52				#DIV/0!	#DIV/0!				
53				#DIV/0!	#DIV/0!				
54				#DIV/0!	#DIV/0!				
55				#DIV/0!	#DIV/0!				
56				#DIV/0!	#DIV/0!				
57				#DIV/0!	#DIV/0!				
58				#DIV/0!	#DIV/0!				
59				#DIV/0!	#DIV/0!				
60				#DIV/0!	#DIV/0!				
61				#DIV/0!	#DIV/0!				
62				#DIV/0!	#DIV/0!				
63				#DIV/0!	#DIV/0!				
64				#DIV/0!	#DIV/0!				
65				#DIV/0!	#DIV/0!				
66				#DIV/0!	#DIV/0!				
67				#DIV/0!	#DIV/0!				
68				#DIV/0!	#DIV/0!				
69				#DIV/0!	#DIV/0!				
70				#DIV/0!	#DIV/0!				
71				#DIV/0!	#DIV/0!				
72				#DIV/0!	#DIV/0!				
73				#DIV/0!	#DIV/0!				
74				#DIV/0!	#DIV/0!				
75				#DIV/0!	#DIV/0!				
76				#DIV/0!	#DIV/0!				
77				#DIV/0!	#DIV/0!				
78				#DIV/0!	#DIV/0!				
79				#DIV/0!	#DIV/0!				
80				#DIV/0!	#DIV/0!				

41				#DIV/0!	#DIV/0!				
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Yfiler

RX#	Sample	Conc ng/ul	Dil	Vol. Ampl. (ul)	TE	Results OK	RI	RI 8 sec	RI 3 sec
81				#DIV/0!	#DIV/0!				
82				#DIV/0!	#DIV/0!				
83				#DIV/0!	#DIV/0!				
84				#DIV/0!	#DIV/0!				
85				#DIV/0!	#DIV/0!				
86				#DIV/0!	#DIV/0!				
87				#DIV/0!	#DIV/0!				
88				#DIV/0!	#DIV/0!				
89				#DIV/0!	#DIV/0!				
90				#DIV/0!	#DIV/0!				
91				#DIV/0!	#DIV/0!				
92				#DIV/0!	#DIV/0!				
93				#DIV/0!	#DIV/0!				
94				#DIV/0!	#DIV/0!				
95				#DIV/0!	#DIV/0!				
96				#DIV/0!	#DIV/0!				

[illegible]

3130 Plate Reagent Calculations

Date: _____

GlobalFiler			
Kit Component	Volume per reaction (µl)		# of samples +extra
Formamide	19.2	X	
GS 600 LIZ	0.8	X	

PP16HS			
Kit Component	Volume per reaction (µl)		# of samples +extra
Formamide	24	X	
ILS 600	1	X	

Yfiler			
Kit Component	Volume per reaction (µl)		# of samples +extra
Formamide	24.5	X	
LIZ	0.5	X	

Amount to pipette
0
0

Amount to pipette
0
0

Amount to pipette
0
0

Date: _____

3130 Plate Set-up

	1	2	3	4	5	6
A						
B						
C						
D						
E						
F						
G						
H						

Analyst: _____

3130 Data File Name: _____

Setup Manually: _____

[illegible]